

# Differential Effects of Haloperidol and Clozapine on Ionotropic Glutamate Receptors in Rats

CHRISTOPHER F. SPURNEY,<sup>1</sup> SERAPIO M. BACA,<sup>1</sup> ANGELA M. MURRAY,<sup>1</sup> GEORGE E. JASKIW,<sup>2</sup>  
JOEL E. KLEINMAN,<sup>1</sup> AND THOMAS M. HYDE<sup>1\*</sup>

<sup>1</sup>NIMH, Clinical Brain Disorders Branch, NIMH, IRP, NIH, Bethesda, Maryland 20892.

<sup>2</sup>Psychiatry Service 116A(B), Cleveland VAMC, Cleveland, Ohio 44141.

**KEY WORDS** glutamate receptors; brain; haloperidol; clozapine; rat

**ABSTRACT** Despite multiple lines of investigation the effect of neuroleptics on glutamate-mediated neurotransmission remains controversial. To study the effects of typical and atypical neuroleptics on selected parameters of glutamate-mediated neurotransmission, male Sprague-Dawley rats were randomly assigned to a 21-day oral treatment course with vehicle, haloperidol (HDL), or clozapine (CLZ). Coronal slices of rat brain were then incubated with tritiated ligands to measure NMDA, AMPA, and kainate receptor, and glutamate reuptake site density. Regions of interest included the frontal cortex, anterior cingulate cortex, dorsal striatum, ventral striatum, and the nucleus accumbens. CLZ increased the density of AMPA receptors significantly in the frontal and anterior cingulate cortices compared with normal controls. In the dorsal and ventral striatum, and nucleus accumbens as a whole, CLZ-treated rats had a higher AMPA receptor density compared with both the HDL- and vehicle-treated controls. Additionally, within the nucleus accumbens, CLZ-treated rats had a higher density of AMPA receptors compared with the HDL group in the core, and at trend level in the shell. There was a group by region interaction for NMDA receptor density, primarily reflecting the tendency of HDL treated rats to have high receptor densities in the frontal and anterior cingulate cortices. Kainate receptors and glutamate reuptake site densities did not differ significantly across groups. These results suggest a critical role for glutamate in the mediation of atypical antipsychotic drug action in anatomically-specific regions, and further encourage the investigation of glutamate neurotransmitter systems in schizophrenia. **Synapse 34:266-276, 1999.** Published 1999 Wiley-Liss, Inc.†

## INTRODUCTION

Neuroleptics are the mainstay of the pharmacological treatment for schizophrenia. The role of glutamate, an excitatory amino acid neurotransmitter, and the effects of neuroleptics on glutamatergic neurotransmission in the pathophysiology of schizophrenia, are controversial topics. Both typical and atypical neuroleptics have significant effects on glutamate-mediated neurotransmission, suggesting a role for glutamate in the pathophysiology of psychosis. Haloperidol (HDL), a typical neuroleptic, is a potent antagonist of the dopamine (DA) D<sub>2</sub> receptor (Seeman, 1980). Activation of D<sub>2</sub> receptors reduces glutamate release in the striatum (Crowder and Bradford, 1987; Mitchell and Daggett, 1980), whereas agonists to the NMDA receptor complex mostly increase DA release (Carter, 1988; Clow and Jahamandas, 1989; Imperato et al., 1990; Krebs et al., 1991). Chronic neuroleptic treatment has also been

implicated in DA supersensitivity, via a decrease in intrastriatal glutamate transmission, which may be partially responsible for the antipsychotic effects of HDL (Freed, 1988). These findings collectively suggest that typical neuroleptics might have significant effects on the glutamate systems that may underlie some of their therapeutic actions.

Clozapine (CLZ) is an atypical but extremely potent neuroleptic effective in treating refractory schizophrenia. In contrast to HDL, CLZ treatment induces relatively few extrapyramidal side effects (Meltzer, 1991). CLZ has an unusual pharmacological profile compared to HDL and other typical neuroleptics (Altar et al., 1986; Goldstein et al., 1989; Huttunen, 1995; Meltzer et

\*Correspondence to: Thomas M. Hyde, M.D., Ph.D., Suite 233, 4701 Willard Avenue, Chevy Chase, Maryland 20815. Telephone No. 301-652-8777; FAX No. 301-652-0856.

Received 13 February 1999; Accepted 19 March 1999

al., 1989; Wiesel et al., 1994). However, the precise characteristics that confer its atypical status remain speculative because CLZ acts on so many neurochemical systems (Deutch et al., 1991; Kerwin, 1994). A number of neurotransmitters, including glutamate, could contribute to CLZ's atypical profile and clinical efficacy (Yamamoto et al., 1994). A modulatory role for CLZ on glutamate neurotransmission can be inferred from its ability to suppress striatal responses from glutamate afferents even after DA depletion (Lidsky and Banerjee, 1992). Moreover, CLZ has been reported to differ from HDL in its effect on glutamate release in the striatum (Yamamoto and Cooperman, 1994). However, both HDL and CLZ can directly displace the binding of tritiated MK-801 from the NMDA receptor (Janowsky and Berger, 1989; Lidsky et al., 1993). The direct effect of CLZ on glutamate receptor density is controversial. Electrophysiologically, CLZ potentiates NMDA-mediated neurotransmission, while HDL depresses the AMPA-mediated response in rat prefrontal cortex (Arvanov et al., 1997). The effect of CLZ on the AMPA-mediated response showed a trend in the same direction. These findings suggest a differential effect of these drugs on central glutamate systems.

The glutamate hypothesis of schizophrenia was inspired by the finding of decreased glutamate levels in the cerebrospinal fluid (CSF) of schizophrenics (Kim et al., 1980). Subsequently, phencyclidine (PCP) was found to cause psychotic symptoms at least in part through NMDA receptor blockade, prompting a second-generation hypothesis of abnormal NMDA receptor function in schizophrenia (Anis et al., 1983). Recent data regarding a glutamate hypothesis has been limited and conflicting. Several studies failed to find decreased CSF levels of glutamate in schizophrenic patients (Gattaz et al., 1985; Korpi et al., 1987), although others found an inverse correlation between CSF glutamate levels and positive symptoms in schizophrenic patients (Faustman et al., 1999). A post-mortem study of glutamate levels did not find significant changes in the frontal cortex, caudate nucleus, putamen, thalamus, nucleus accumbens, or olfactory tubercle (Perry, 1982). Nevertheless, the psychotogenic effects of NMDA antagonists are well established, suggesting that glutamate may play a role in the generation of psychotic symptoms in schizophrenia.

Post-mortem studies of glutamate receptors in schizophrenia vary by region and receptor sub-type. Two reports noted increased kainate receptor density in the medial frontal and "eye movement" frontal cortices (Nishikawa et al., 1983; Toru et al., 1988). Another found increased kainate receptor density in the orbital frontal cortex of schizophrenics (Deakin et al., 1989). In separate studies, Kerwin and colleagues reported decreased kainate receptor levels in the hippocampus and parahippocampal gyrus (Kerwin et al., 1988, 1990). Increased levels of NMDA NR2 receptor subunit mRNA

expression have been demonstrated in the prefrontal cortex of schizophrenics, suggesting that the frontal lobe deficits seen in schizophrenia may be related to decreased glutamate neurotransmission or receptor function (Akbarian et al., 1996). In 1989, Kornhuber and colleagues found a significant increase in NMDA receptor density in the putamen, among other regions (Kornhuber et al., 1989). This finding has been bolstered by a report of deficient glutamate reuptake sites in the basal ganglia (Simpson et al., 1992). Noga and colleagues did not find changes in basal ganglia NMDA or kainate receptor density of schizophrenic patients, but reported a significant increase in AMPA receptor density in the ventral caudate (Noga et al., 1997). No consensus has emerged regarding the site or type of glutamate receptor changes associated with schizophrenia. More importantly, the changes noted in post-mortem tissue may be secondary to neuroleptics rather than reflecting the primary pathology of schizophrenia.

Despite the notion that glutamate systems in general, and NMDA receptors in particular, may be involved in psychosis, the effects of the antipsychotics on ionotropic glutamate receptor systems have received little attention until recently. Several studies have examined the effects of neuroleptics on glutamate receptor systems, but a clear consensus is lacking (Fitzgerald et al., 1995; Hamid et al., 1998; Healy and Meador-Woodruff, 1997; Kakigi et al., 1992; McCoy et al., 1998; Oretti et al., 1994; Tarazi et al., 1996). To further elucidate the involvement of glutamate receptor systems in both typical and atypical neuroleptic actions, we investigated the effects of subacute HDL or CLZ treatment on glutamate ionotropic receptor densities and glutamate reuptake sites in selected regions of rat brain. Regions were selected for their putative involvement in the pathophysiology of psychosis. Antipsychotic drugs were delivered in drinking water at doses chosen to approximate those used clinically. We postulated that subacute CLZ and HDL treatments would differentially affect glutamate receptors due to their separate pharmacological profiles.

## MATERIALS AND METHODS

### Material

The ligands [vinylidene-<sup>3</sup>H]-kainic acid (58.0 Ci/mmol), d-[2,3-<sup>3</sup>H]-aspartic acid (11.5 Ci/mmol), (+)-[3-<sup>3</sup>H]-MK-801 (20.3 Ci/mmol) and [5-<sup>3</sup>H]-CNQX (26.7 Ci/mmol) were obtained from Dupont-NEN (Boston, MA). Kainate and d-aspartic acid were obtained from Research Biochemicals International (Natick, MA). L-Glutamate, ketamine and all other chemicals were provided by Sigma Chemical Company (St. Louis, MO).

### Treatment groups

Male Sprague-Dawley rats (Zivic-Miller), 200–250 g, were housed 3/cage with food and water ad libitum on a

standard 12-hour dark/light cycle. Animals were randomly assigned to a 21-day treatment course with vehicle (VEH), haloperidol (HDL) or clozapine (CLZ). Antipsychotic drugs were delivered in drinking water. Clozapine has a very short half-life in rodents, and single daily intraperitoneal injections do not result in sustained levels of clozapine in the brain (Baldessarini et al., 1993). Animals received either VEH (dilute tartaric acid buffered to pH 7.3 with NaHCO<sub>3</sub>), HDL (1 mg/kg/d) or CLZ (20 mg/kg/d). Both HDL and CLZ were first dissolved in 0.1 M tartaric acid, brought to volume with distilled water and buffered to pH 7.1–7.3 with 1 M NaHCO<sub>3</sub>. Solutions were replenished every two days.

This experiment utilized three groups of rats: Group (1)—rats treated with VEH (n = 11), Group (2)—rats treated 21 days with HDL (n = 6), and Group (3)—rats treated 21 days with CLZ (n = 8). All procedures involving animal use were in strict compliance with the NIH Guide for the Care and Use of Laboratory Animals.

### Preparation of brain tissue

After a 72-hour period of drug withdrawal, animals were sacrificed without anesthesia. Their brains were removed rapidly, quick-frozen on dry ice, and stored at –70°C. Twenty-micron thick coronal sections were cut rostrocaudally using a cryostat (Jung Frigocut, Nussloch, Germany) and thaw-mounted onto acid-scrubbed, double-subbed, gelatin-coated slides. Sections were taken at the level of the anterior commissure. Slide mounted sections were stored in slide boxes sealed in plastic at –70°C until used in assays.

### Quantitative receptor autoradiography

Before experiments, all sections were thoroughly blown dry with cool air for 30 minutes. After experiments, they were blown dry with cool air for 3 minutes.

### CNQX

Preincubation consisted of 15 minutes at 4°C followed by 15 minutes at 30°C in a 50 mM Tris-HCl buffer at pH 7.2. Sections were then incubated for 45 minutes at 4°C in buffer containing 50 nM <sup>3</sup>H-CNQX. Non-specific binding was determined using 1 mM L-glutamic acid (modified from Kerwin et al., 1988). Sections were then rapidly dipped three times in ice cold 50 mM Tris-HCl buffer.

### MK-801

Sections were incubated for 150 minutes in a buffer containing 30 mM EPPS, 100 μM L-glutamate, 100 μM L-glycine, and 1 mM EDTA (pH 7.45) at room temperature. Total binding was determined by the addition of 16 nM <sup>3</sup>H-MK-801 and non-specific binding with the addition of 200 μM ketamine. Sections were then washed twice in fresh ice cold buffer that did not contain either glutamate or glycine for a total of 40

minutes (modified from Subramaniam and McGonigle, 1991).

### Kainic acid

Preincubation consisted of 15 minutes at 4°C followed by 15 minutes at 30°C in a 50 mM Tris-HCl buffer at pH 7.0. Incubation occurred for 120 minutes at 4°C in buffer containing 24 nM <sup>3</sup>H-kainic acid. Non-specific binding was determined by the addition of 100 μM kainate. The sections were then rapidly dipped three times in fresh ice cold 50 mM Tris-HCl buffer with a final dip in ice cold distilled water (McGonigle, personal communication).

### D-aspartate

Preincubation consisted of 10 minutes at 30°C in a 50 mM Tris HCl buffer containing 300 mM NaCl at pH 7.4. Sections were then incubated for 20 minutes at 4°C in a buffer containing 100 nM <sup>3</sup>H-d-aspartic acid. Non-specific binding was determined using 100 μM cold d-Aspartic acid (modified from Anderson et al., 1990). Sections were rinsed four times for a total of 30 seconds in ice cold 50 mM Tris-HCl and 300 mM NaCl buffer.

### Image generation and analysis

All sections were placed in boxes along with desiccant overnight before being placed into cassettes with tritiated microscales (Amersham, Arlington Heights, IL) and apposed to [<sup>3</sup>H] Hyperfilm (LKB Instruments, Gaithersburg, MD). Exposure times were four days for CNQX and kainic acid, 15 days for d-aspartate and 13 days for MK-801. Films were developed and scanned into a Macintosh computer (Apple Computers, Cupertino, CA). Receptor densities were measured using an image analysis system (NIH Image 1.52, Wayne Rasband, RSB, NIMH).

Quantitative densitometry was used to measure receptor density in the frontal cortex, anterior cingulate cortex, striatum, and nucleus accumbens. The striatum was subdivided into dorsal and ventral regions, while the nucleus accumbens was subdivided into the core and shell [see Fig. 1]. Each slide contained two sections and the four measured areas from the films were averaged. Specific binding was determined by subtracting nonspecific binding from the total binding.

### Statistics

Comparisons between groups were carried out using a repeated measures analysis of variance (rmANOVA) for each ligand, to look for a main effect of group, and the effects of region, and group by region interactions. If there was a main effect of group ( $p < 0.05$ ), a one-way ANOVA was conducted for each brain region, followed by post-hoc Fisher's PLSD t-tests.

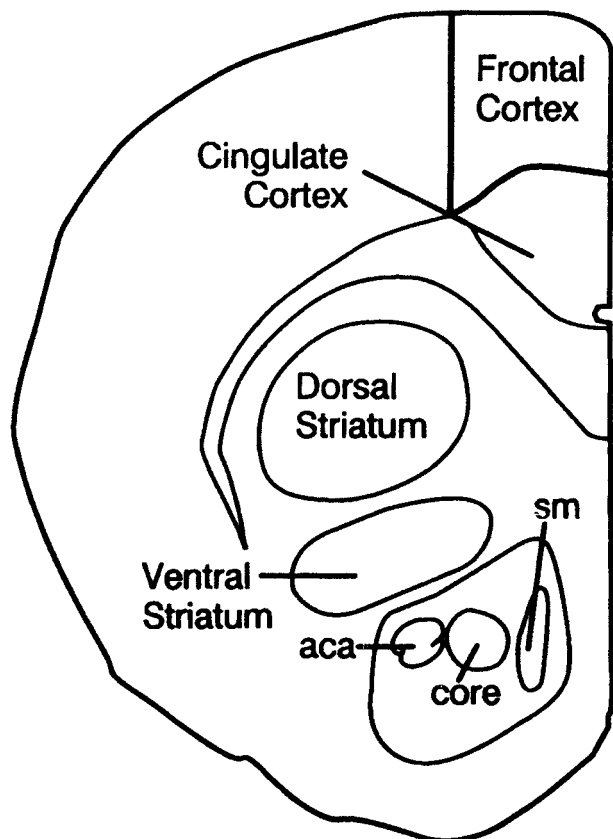


Fig. 1. Schematic of the regions measured in the rat brain to assess receptor binding levels.

## RESULTS

### AMPA receptors

By rmANOVA, there was a main effect of region ( $df = 5, 110; F = 76.1; p < .001$ ). CNQX binding was highest in the anterior cingulate cortex (679 fm/mg), followed by the frontal cortex (644 fm/mg), accumbens-core (514 fm/mg), ventral striatum (505 fm/mg), accumbens-total (455 fm/mg), dorsal striatum (407 fm/mg), and accumbens-shell (402 fm/mg) (see Fig. 2).

Analysis of CNQX binding was also significant for a main effect of group ( $df = 2, 22; F = 4.35; p = .026$ ) [see Fig. 3]. ANOVA analyses for group differences were significant in the frontal cortex ( $p = .037$ ), anterior cingulate cortex ( $p = .02$ ), dorsal striatum ( $p = .007$ ), ventral striatum ( $p = .014$ ), and nucleus accumbens as a whole ( $p = .027$ ). Post-hoc t-tests revealed significantly higher AMPA receptor density in the frontal cortex and anterior cingulate of CLZ-treated rats relative to vehicle treated controls ( $p = .011$  and  $.006$ , respectively). However, in the dorsal striatum, ventral striatum, and nucleus accumbens as a whole, CLZ-treated rats had a higher AMPA receptor density compared to both the HDL- and vehicle-treated controls ( $p < .05$ ). Within the nucleus accumbens, CLZ-treated rats differed significantly from the HDL group in the

core ( $p = .02$ ), and at trend level in the shell ( $p = .06$ ). Finally, there was a trend toward decreased AMPA receptor density in the accumbens shell following HDL treatment ( $p = .07$ ).

There was also a group by region interaction ( $df = 10, 110; F = 3.02; p = .002$ ); this interaction may reflect the consistently increased density of AMPA receptors across both cortical and subcortical structures following CLZ treatment, and a decreased density following HDL [see Fig. 4].

### NMDA receptors

By rmANOVA, there was a main effect of region ( $df = 5, 110; F = 58.4; p < .001$ ). MK-801 binding was highest in the anterior cingulate cortex (470 fm/mg), followed by the frontal cortex (354 fm/mg), ventral striatum (249 fm/mg), accumbens-total (222 fm/mg), accumbens-shell (209 fm/mg) and accumbens-core (202 fm/mg). There was no main effect of group, but there was a group by region interaction ( $df = 10, 110; F = 2.46; p < .01$ ) [see Fig. 5]. This interaction may reflect the higher densities of NMDA receptors in cortical regions following HDL treatment compared to CLZ, as well as relatively equal binding densities in subcortical structures.

### Kainic acid receptors

By rmANOVA, there was a main effect of region ( $df = 5, 110; F = 173.0; p < .001$ ). Kainic acid binding was highest in the ventral striatum (199 fm/mg), followed by the dorsal striatum (172 fm/mg), accumbens-core (161 fm/mg), accumbens-total (152 fm/mg), anterior cingulate cortex (148 fm/mg), accumbens-shell (135 fm/mg) and frontal cortex (131 fm/mg) (see Fig. 2). No significant differences were seen between the groups, nor was there any group by region interaction (see Table 1).

### Glutamate reuptake sites

By rmANOVA, there was a main effect of region ( $df = 5, 110; F = 27.8; p < .001$ ). D-aspartic acid binding was highest in the anterior cingulate cortex (869 fm/mg) followed by the frontal cortex (853 fm/mg), ventral striatum (758 fm/mg), accumbens-core (752 fm/mg), dorsal striatum (708 fm/mg), accumbens-total (694 fm/mg), and accumbens-shell (657 fm/mg) (see Fig. 2). No significant differences were seen between the groups, nor was there any group by region interaction (see Table 1).

## DISCUSSION

We examined the density of ionotropic glutamate receptors and reuptake sites in selected brain regions of normal rats treated for 21 days with orally administered HDL, CLZ, or vehicle. CLZ significantly increased the density of AMPA receptors in the frontal and

### Regional Receptor Density in Normal Rats

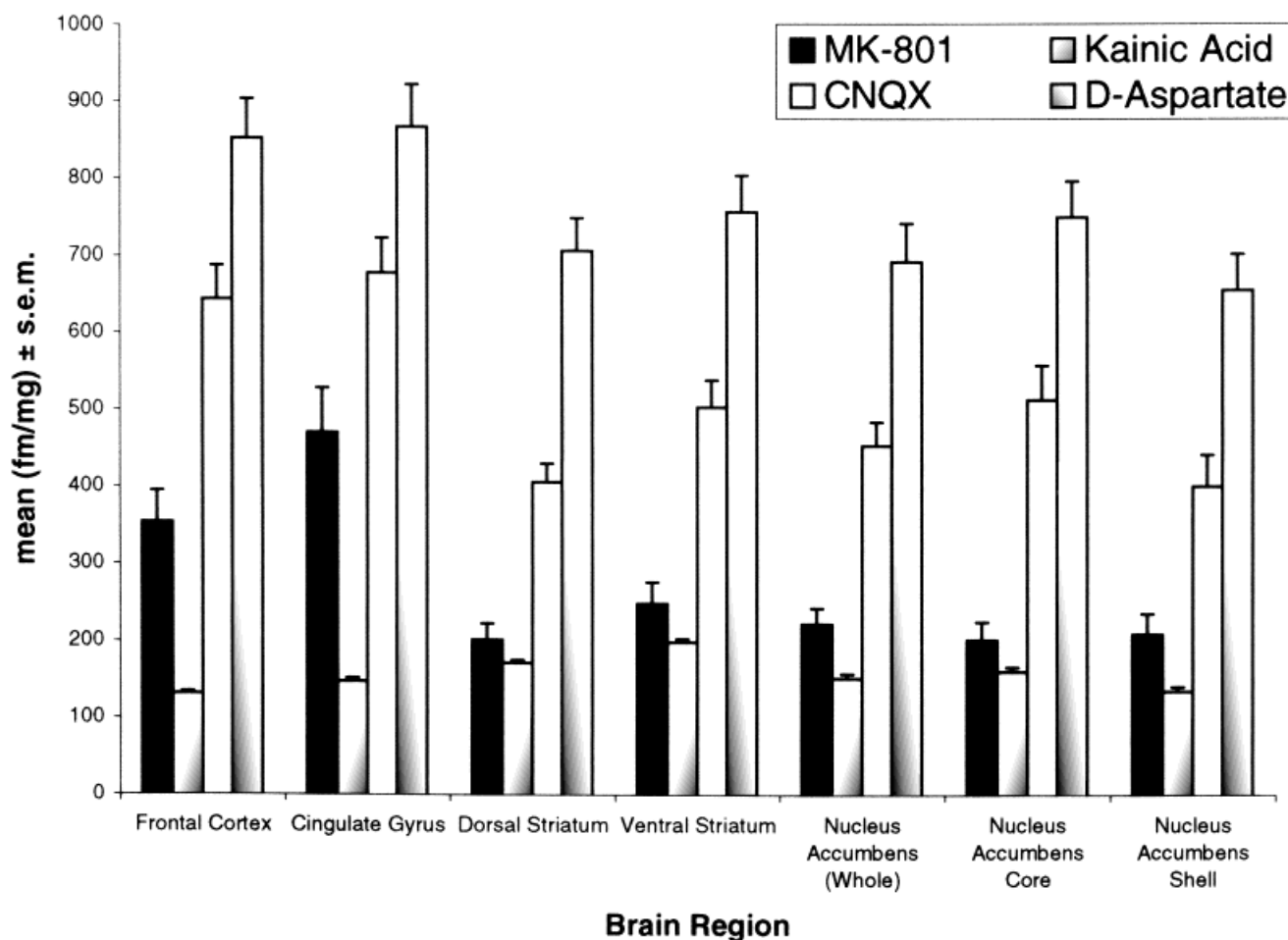


Fig. 2. Relative distribution of receptor density for NMDA, AMPA, kainate, and reuptake sites in vehicle treated control rats.

anterior cingulate cortices compared with normal controls. Moreover, in the dorsal striatum, ventral striatum, and nucleus accumbens as a whole, CLZ-treated rats had a higher AMPA receptor density compared to both the HDL- and vehicle-treated controls. Within the nucleus accumbens, CLZ-treated rats had a higher density of AMPA receptors compared to the HDL group in the core, and achieved trend level in the shell. There was a group by region interaction for NMDA receptor density, primarily reflecting the tendency of HDL-treated rats to have high receptor densities in the frontal and anterior cingulate cortices. Kainate receptors and glutamate reuptake site densities did not differ significantly across groups. These findings suggest a differential effect of CLZ and HDL on glutamate neurotransmission in both cortical and subcortical regions. The unique efficacy of CLZ may be derived, at least in part, from these differential effects.

There is no consensus regarding the effects of neuroleptic treatment on AMPA receptors. McCoy et al., 1998 found that HDL (0.5 mg/kg/day i.p. for 21 days) but not CLZ (20 mg/kg/day i.p. for 21 days) increased the density of AMPA receptors in the frontal cortex and the dorsal and ventral striatum. Tarazi et al., 1996 did not find significant changes in AMPA receptor density after treatment with HDL (1.5 mg/kg/day p.o. for 28 days or 8 months) or CLZ (25 mg/kg/day p.o. for 28 days or 8 months) (Tarazi et al., 1996). Kakigi and colleagues noted that chronic HDL treatment (6 months at 1.5 mg/kg/day p.o.) decreased AMPA receptor density in the nucleus accumbens, which our data tend to support (Kakigi et al., 1992). Clearly there is considerable dispute about the effects of neuroleptics on AMPA receptors.

AMPA receptors are aggregates of four types of subunits, designated GluR1–4. In support of our findings, CLZ appears to increase GluR2 mRNA expression

### CNQX Receptor Density following Haloperidol, Clozapine, and Vehicle Treatment

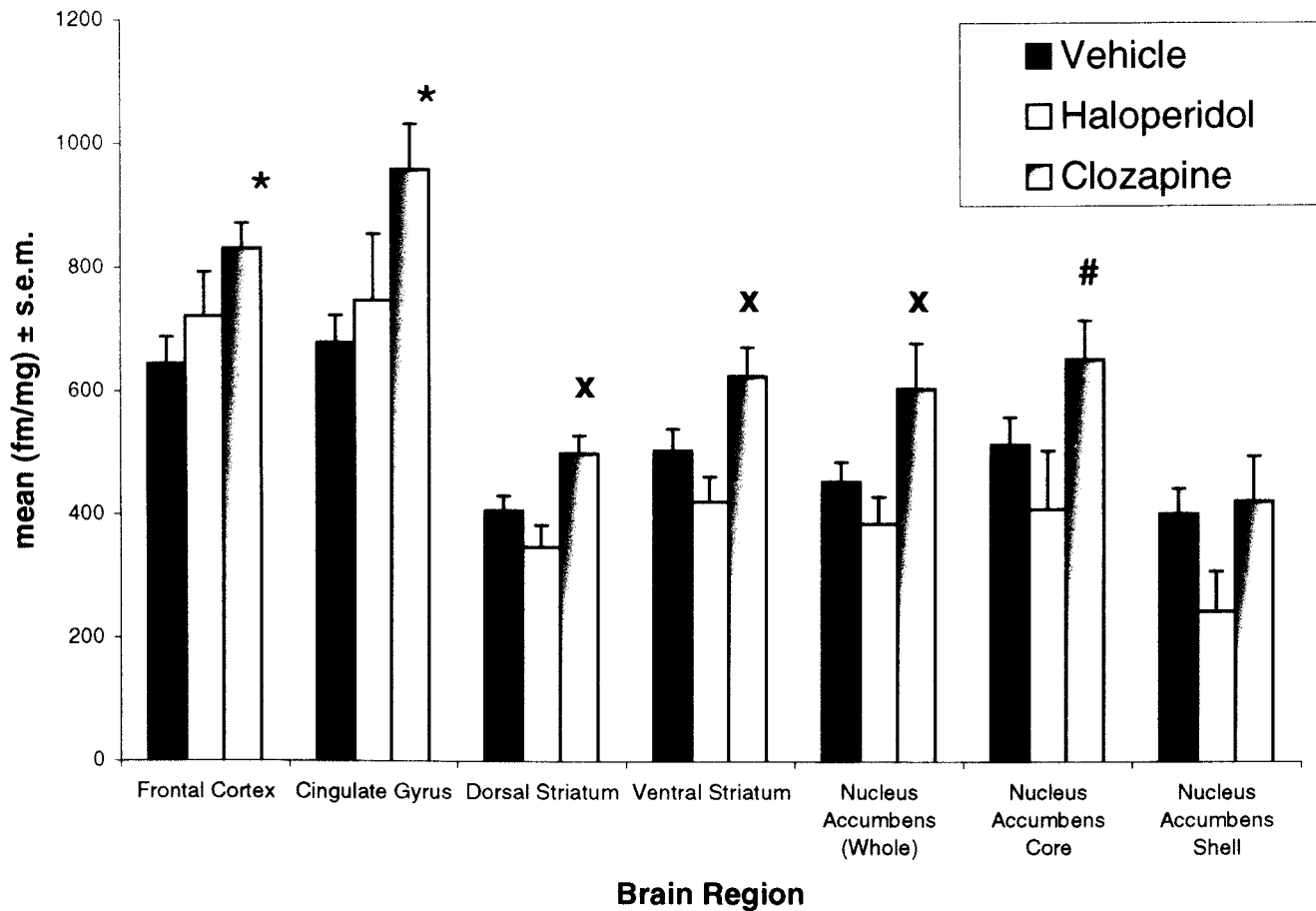


Fig. 3. AMPA receptor density was higher following CLZ treatment compared to vehicle treated controls in frontal and anterior cingulate cortices ( $p < .05$ ) (\*). In the dorsal and ventral striatum and nucleus accumbens, CLZ treated rats had a higher density of AMPA

receptors compared to both HDL and vehicle-treated groups ( $p < .05$ ) (X). In the core region of the nucleus accumbens, CLZ treated rats had a higher density than the HDL group ( $p < .05$ ) (#).

in the frontal/parietal cortex, nucleus accumbens, and hippocampus (Fitzgerald, 1995), although this finding has been contested (McCoy et al., 1998; Oretti, 1994). Healy and Meador-Woodruff (1997) found that HDL (2 mg/kg/day s.c. for 14 days) actually decreased GluR2 and GluR4 mRNA expression in the frontal cortex and striatum, which our data tend to support. Counter to our report, Healy and Meador-Woodruff (1997) also reported that CLZ (20 mg/kg/day S.C. for 14 days) produced a decrease in GluR3 mRNA expression in the frontal cortex and striatum, and a decrease in GluR4 mRNA expression in the striatum, however, there is not always a direct correlation between receptor mRNA expression and protein levels in the central nervous system. Additionally, the subunit composition of the AMPA receptor might be altered by neuroleptic treatment without effecting the overall receptor density.

The conflicting results from studies of AMPA receptors following neuroleptic treatment may be secondary to methodological considerations. Differences in dose, route of administration, and duration of treatment must be considered. Oretti et al. (1994) administered HDL 1.5 mg/kg/day i.p., while McCoy et al. (1998) used HDL at a dose of 0.5 mg/kg/day i.p. or CLZ at 20 mg/kg/day i.p., for 21 days. Since HDL is subject to first-pass metabolism, after injection HDL levels are considerably higher than the same-but-orally-administered doses (Bianchetti et al., 1980). Other groups have used higher HDL doses: 2.0 mg/kg/day (Healy and Woodruff, 1997), 1.8 mg/kg/day (Fitzgerald et al., 1995) and 1.5 mg/kg/day (Kakigi et al., 1992; Tarazi et al., 1996), and CLZ doses: 35 mg/kg/day (Fitzgerald et al., 1995) and 25 mg/kg/day (Tarazi et al., 1996) than we employed. Differences in treatment regimens may un-

### AMPA Receptor Density by Brain Region and Group

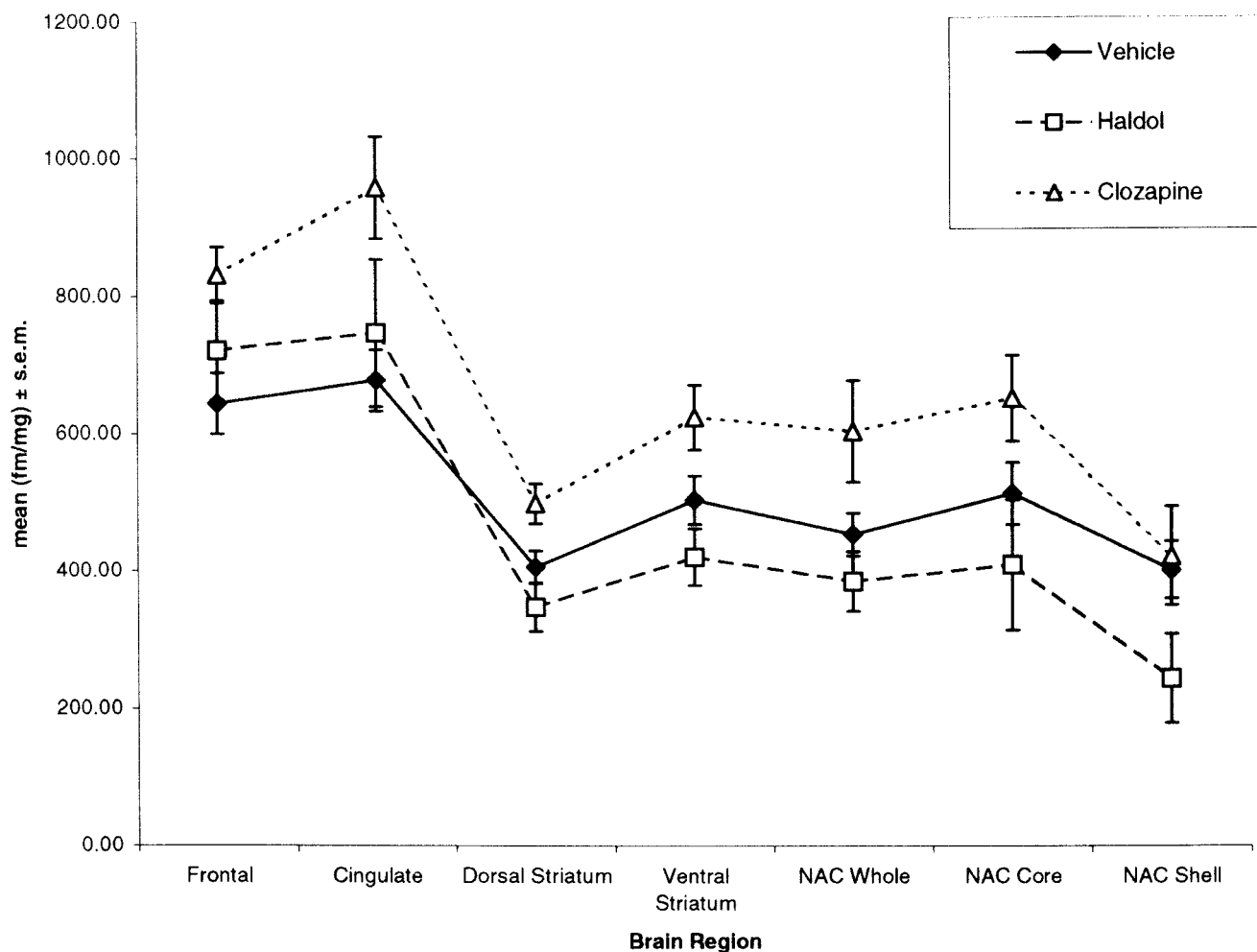


Fig. 4. Graphic representation of the binding density of AMPA receptors across selected brain regions following CLZ, HDL, and vehicle treatment. The group by region interaction is reflected in the

relatively high density of AMPA receptors in the cortical regions following CLZ treatment, and the trend towards decreased basal ganglia receptor density following HDL treatment.

derlie the discrepancies between studies of glutamate receptor density and subunit mRNA expression levels.

Since compounds that disrupt NMDA receptor function induce psychotic symptoms, the mechanism of action of neuroleptics might involve an increase in the density of NMDA receptors. However, the absence of a significant group effect on NMDA receptor density in our study is consistent with the report of Meshul et al. (1996) and a previous report from our laboratory (Hamid, 1998). Meshul et al. (1996) did not find a change in the density of NMDA receptors in the striatum after a 28-day treatment with either HDL (0.5 mg/kg/day s.c.) or CLZ (30 mg/kg/day s.c.). We found no change in the striatum after HDL treatment (1 mg/kg/day i.p. for 21 days). In a study of Tarazi et al. (1996), treatment with CLZ (25 mg/kg/day p.o. for 28 days) actually caused a reduction of NMDA receptor density

in the striatum. Giardino et al., 1997 treated rats for 21 days with CLZ (30 mg/kg/day i.p.), and found unilateral decreases in NMDA receptor density in the left anterior cingulate, frontoparietal motor, and frontoparietal somatosensory cortices. They did not detect changes in the prefrontal cortex, striatum, or nucleus accumbens. We did not fractionate our data by hemisphere, preventing direct comparison with their cortical findings; however, our subcortical results are largely in agreement. While there was a tendency for CLZ to reduce cortical NMDA receptor density in our study, it did not reach statistical significance. Larger cohorts may have revealed a difference. Although there is extensive data regarding NMDA receptor subunit mRNA expression following neuroleptic treatment, no consensus has emerged (Fitzgerald et al., 1995; Meshul et al., 1996; Oretti et al., 1994; Riva et al., 1997). Neuroleptic-

## NMDA Receptor Density by Brain Region and Group

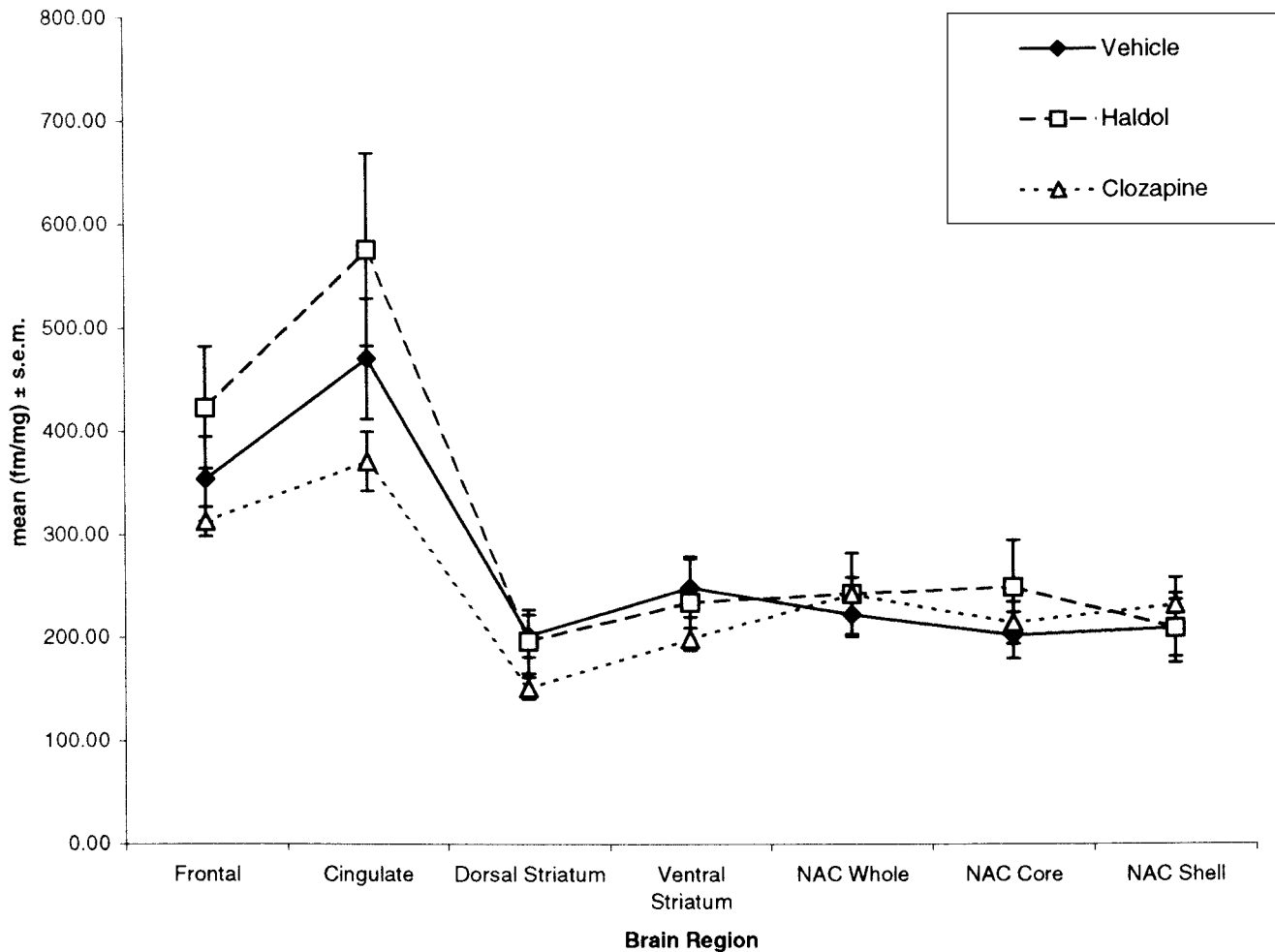


Fig. 5. Graphic representation of the binding density of NMDA receptors across selected brain regions following CLZ, HDL, and vehicle treatment. The group by region interaction is reflected in the relatively high density of NMDA receptors in the cortical regions following CLZ treatment.

induced reductions in NMDA receptor density seem somewhat counter-intuitive given the clinical data regarding NMDA receptor antagonism and psychosis.

Neuroleptics may alter the extracellular levels of glutamate, thereby secondarily altering post-synaptic receptor density. Yamamoto and Cooperman (1994) found 21 days of treatment with CLZ (20 mg/kg/day i.p.) did not produce changes in any brain region while HDL (0.5 mg/kg/day i.p.) increased extracellular glutamate in the caudate and nucleus accumbens (Yamamoto and Cooperman, 1994). In another experiment, Yamamoto and colleagues did not find a change in striatal glutamate levels after 21 days of treatment with CLZ (20 mg/kg/day i.p.), but a five fold increase after 21 days of treatment with HDL (0.5 mg/kg/day i.p.) (Yamamoto et al., 1994). In earlier studies, Bardgett et al. (1993) found no differences in glutamate levels in the striatum

after 28 days of treatment with HDL (0.18 mg/kg/day s.c.) or CLZ (19.4 mg/kg/day s.c.). However, extracellular glutamate levels may not accurately reflect the amount of glutamate released from axon terminals since changes may reflect alterations in metabolism or GABA-mediated transmission rather than selective changes in the neurotransmitter pool (Deutch et al., 1989).

Neuroleptics might effect glutamate neurotransmission through indirect mechanisms. Dopaminergic nigrostriatal fibers may mediate a presynaptic inhibitory influence via  $D_2$  receptors located on glutamatergic corticostriatal terminals in the striatum (Kim et al., 1980; Kornhuber and Kornhuber, 1986). Thus, a hyperdopaminergic state such as schizophrenia might lead to decreased levels of glutamate release in the striatum. In such a case, neuroleptic-induced  $D_2$  receptor block-



TABLE I. Receptor binding results (mean  $\pm$  s.e.m. fm/mg) for MK-801, kainic acid, CNQX, and *d*-aspartic acid for the frontal cortex, cingulate gyrus, dorsal striatum, ventral striatum, nucleus accumbens-total, nucleus accumbens-core, and nucleus accumbens-shell

Ligand	Group	Frontal cortex	Cingulate gyrus	Dorsal striatum	Ventral striatum	N accumbens—total	N. accumbens—core	N. accumbens—shell
MK-801	Normal	354.18 $\pm$ 41	470.43 $\pm$ 58	202.06 $\pm$ 21	248.57 $\pm$ 28	222.34 $\pm$ 21	202.14 $\pm$ 23	209.00 $\pm$ 27
	HDL	423.18 $\pm$ 59	575.77 $\pm$ 93	196.59 $\pm$ 31	234.71 $\pm$ 44	242.80 $\pm$ 40	248.79 $\pm$ 45	208.88 $\pm$ 33
	CLZ	312.87 $\pm$ 14	371.07 $\pm$ 29	151.26 $\pm$ 11	198.90 $\pm$ 11	241.98 $\pm$ 17	214.41 $\pm$ 20	230.97 $\pm$ 27
Kainic Acid	Normal	131.15 $\pm$ 3	147.86 $\pm$ 4	171.74 $\pm$ 4	199.25 $\pm$ 4	151.90 $\pm$ 6	160.77 $\pm$ 6	134.79 $\pm$ 6
	HDL	140.05 $\pm$ 7	149.59 $\pm$ 7	173.16 $\pm$ 9	201.24 $\pm$ 9	152.54 $\pm$ 13	163.32 $\pm$ 14	137.09 $\pm$ 6
	CLZ	128.21 $\pm$ 5	152.21 $\pm$ 7	169.91 $\pm$ 6	203.27 $\pm$ 7	146.13 $\pm$ 6	157.93 $\pm$ 4	131.02 $\pm$ 6
CNQX	Normal	644.24 $\pm$ 44	678.71 $\pm$ 45	406.94 $\pm$ 24	504.65 $\pm$ 35	454.67 $\pm$ 31	513.88 $\pm$ 45	401.95 $\pm$ 41
	HDL	721.60 $\pm$ 72	747.66 $\pm$ 108	348.53 $\pm$ 35	421.78 $\pm$ 41	386.10 $\pm$ 44	409.78 $\pm$ 95	244.03 $\pm$ 65
	CLZ	831.67 $\pm$ 41*	959.74 $\pm$ 74*	499.46 $\pm$ 29**	625.38 $\pm$ 47**	605.07 $\pm$ 74**	652.72 $\pm$ 62***	422.83 $\pm$ 72
<i>D</i> -aspartate	Normal	853.24 $\pm$ 51	868.63 $\pm$ 55	707.67 $\pm$ 43	758.45 $\pm$ 48	693.67 $\pm$ 50	752.17 $\pm$ 47	657.33 $\pm$ 47
	HDL	818.21 $\pm$ 47	811.06 $\pm$ 60	679.87 $\pm$ 34	725.18 $\pm$ 44	628.71 $\pm$ 62	711.38 $\pm$ 63	603.36 $\pm$ 64
	CLZ	699.68 $\pm$ 55	741.94 $\pm$ 67	626.90 $\pm$ 45	683.99 $\pm$ 46	625.76 $\pm$ 41	723.45 $\pm$ 57	577.22 $\pm$ 43

\*Significant ( $p < .05$ ) difference when compared to control group.

\*\*Significant ( $p < .05$ ) difference when compared to HDL and control groups.

\*\*\*Significant ( $p < .05$ ) difference when compared to HDL group.

ade might decrease dopamine, restore glutamate and thus produce antipsychotic effects. In normal rats, neuroleptics may decrease striatal dopamine release, resulting in relatively excessive amounts of glutamate release. In turn, this may cause a down-regulation in some post-synaptic glutamate receptors or changes in glutamate receptor subunit composition. The cortico-thalamocortical negative feedback loop also may play a role in neuroleptic-induced glutamate receptor density changes (Carlsson and Carlsson, 1990). In this model, corticostriatal excitatory glutamatergic and nigrostriatal inhibitory dopaminergic systems independently modulate GABA-mediated inhibition of the excitatory thalamocortical pathway. Increased dopamine might decrease the inhibitory GABA-mediated signal from the basal ganglia to the thalamus, and putatively decreased glutamate release could lead to further disinhibition of the thalamocortical signals, in turn eliciting the clinical symptoms of hyperarousal, disturbed perception, and psychotic behavior (Wachtel and Turski, 1990).  $D_2$  blockade would ameliorate symptoms in such a model by attenuating the DA mediated inhibition of GABA transmission from the basal ganglia to the thalamus. Our experiment found negligible changes in glutamate receptor density following HDL treatment, potentially supporting a more independent relationship between glutamate and dopamine as in the cortico-thalamocortical loop, and less of a reciprocal, presynaptically mediated relationship as posited by the initial hypotheses.

CLZ, an atypical neuroleptic, has a pharmacological profile that includes relatively low  $D_2$  affinity (Farde et al., 1988), and high affinity for  $D_4$  receptors (Van Tol, 1991). It also has potent effects on several 5-HT receptor subtypes including 5-HT<sub>2a</sub>, 5-HT<sub>2c</sub>, and 5-HT<sub>3</sub> (Breier, 1995), and antagonism of the histaminergic, cholinergic, and noradrenergic systems (Breier et al., 1994; McMillen and Shore, 1978; Meltzer, 1991; Peroutka et al., 1977; Peroutka and Synder, 1980). CLZ's proposed mechanism of action may involve its effects on

the relative balance between 5-HT and DA systems (Breier, 1995; Meltzer et al., 1989). However, this profile of 5-HT<sub>2</sub>/ $D_2$  antagonism may not explain all of CLZ's effects. In a study of non-human primates, Casey and colleagues found that the addition of a wide range of 5-HT<sub>2</sub> antagonists to typical neuroleptic treatment did not have a potent anti-parkinsonian effect (Casey et al., 1993). Similarly, depletion of central 5-HT did not alter CLZ catalepsy nor did it change the profile of supersensitivity to the DA agonist apomorphine that emerges after chronic CLZ treatment (Jaskiw, personal communication). These results suggest that 5-HT<sub>2</sub> antagonism may not be the sole main mechanism of action for atypical neuroleptics, and the effects on AMPA receptor-mediated neurotransmission may be pertinent.

CLZ may exert some of its unique therapeutic effects through alterations in glutamate neurotransmission. Electrophysiological studies by Lidsky and colleagues show that CLZ has glutamate antagonistic properties which potentially occur at therapeutic levels (Lidsky et al., 1993). Glutamate receptor antagonism may occur primarily through direct antagonism at the NMDA receptor or secondarily through CLZ's antidopaminergic effects. In rat studies, NMDA receptor antagonism reduced catalepsy induced by typical neuroleptics (Elliott et al., 1990; Yoshida et al., 1991), suggesting that CLZ's potential NMDA antagonism may play a role in preventing extrapyramidal side effects. Glutamate has also been shown to increase the release of DA in the rat striatum (Roberts, 1978) and increase burst firing in the ventral tegmental area (Charley et al., 1991). Thus, glutamate antagonism could potentially decrease DA levels and contribute to the antipsychotic profiles of CLZ.

We found minimal effects of HDL or CLZ treatment on NMDA receptor density. This might suggest that as assayed, the NMDA receptors are relatively insensitive to antipsychotic drugs and are not likely to be critically involved in either typical or atypical antipsychotic drug

effects. It is also possible that given the widespread distribution of glutamate and, presumably, glutamatergic receptors; HDL and CLZ only affect certain populations of glutamate receptors. Recent studies suggest that HDL may bind to a site on the NR1A/NR2B subtype of the NMDA receptor (Coughenour and Cordon, 1997). Regions with high density of NMDA receptors of the NR1A/NR2B subtype might be effected differentially by treatment with HDL. Additional investigations need to be performed with CLZ. In addition, regional selectivity could be secondary to the type and amount of dopaminergic innervation of a region. This putative selectivity is noted with significant increases in AMPA receptors in the prefrontal and cingulate cortices. Fitzgerald and colleagues found similar molecular results and proposed a hypothesis of increased AMPA receptor subunits on prefrontal cortex interneurons, potentially increasing GABA-mediated inhibition of glutamatergic output to the subcortical dopaminergic systems (Fitzgerald et al., 1995).

In conclusion, CLZ increased the density of AMPA receptors significantly in the frontal and anterior cingulate cortices compared with normal controls. In the dorsal and ventral striatum, and nucleus accumbens as a whole, CLZ-treated rats had a higher AMPA receptor density compared to both the HDL- and vehicle-treated groups. In the core of the nucleus accumbens, CLZ-treated rats had a higher density of AMPA receptors compared to the HDL group. There was a group by region interaction for NMDA receptor density, primarily reflecting the tendency of HDL-treated rats to have high receptor densities in the frontal and anterior cingulate cortices. Kainate receptors and glutamate reuptake site densities did not differ significantly when HDL and CLZ groups were compared to controls. These results support the notion that alterations in glutamate neurotransmission play a role in the unique therapeutic efficacy of CLZ.

#### ACKNOWLEDGMENTS

The authors thank Terry E. Goldberg, Ph.D. for his valuable advice on statistics.

#### REFERENCES

- Akbarian S, Sucher NJ, Bradley D, Tafazzoli A, Trinh D, Hetrick WP, Potkin SG, Sandman CA, Bunney WE Jr, Jones EG. 1996. Selective alterations in gene expression for NMDA receptor subunits in prefrontal cortex of schizophrenics. *J Neurosci* 16:19–30.
- Altar CA, Wasley AM, Neale RF, Stone GA. 1986. Typical and atypical antipsychotic occupancy of D2 and S2 receptors: an autoradiographic analysis in rat brain. *Brain Res Bull* 16:517–25.
- Anderson KJ, Monaghan DT, Bridges RJ, Tavoularis AL, Cotman CW. 1990. Autoradiographic characterization of putative excitatory amino acid transport sites. *Neuroscience* 38:311–22.
- Anis NA, Berry SC, Burton NR, Lodge D. 1983. The dissociative anaesthetics, ketamine and phencyclidine, selectively reduce excitation of central mammalian neurones by N-methyl-aspartate. *Br J Pharmacol* 79:565–75.
- Arvanov VL, Liang X, Schwartz J, Grossman S, Wang RY. 1997. Clozapine and haloperidol modulate N-methyl-D-aspartate- and non-N-methyl-D-aspartate receptor-mediated neurotransmission in rat prefrontal cortical neurons in vitro. *J Pharmacol Exp Ther* 283:226–34.
- Baldessarini RJ, Centorrino F, Flood JG, Volpicelli SA, Huston-Lyons D, Cohen BM. 1993. Tissue concentrations of clozapine and its metabolites in the rat. *Neuropsychopharm* 9:117–24.
- Bardgett ME, Wrona CT, Newcomer JW, Csernansky JG. 1993. Subcortical excitatory amino acid levels after acute and subchronic administration of typical and atypical neuroleptics. *Eur J Pharmacol* 230:245–50.
- Bianchetti G, Zarifian E, Poirier-Littre MF, Morselli PL, Deniker P. 1980. Influence of route of administration on haloperidol plasma levels in psychotic patients. *Int J Clin Pharmacol Ther Toxicol* 18:324–7.
- Breier A. 1995. Serotonin, schizophrenia and antipsychotic drug action. *Schizophr Res* 14:187–202.
- Breier A, Buchanan RW, Waltrip RW 2nd, Listwak S, Holmes C, Goldstein DS. 1994. The effect of clozapine on plasma norepinephrine: relationship to clinical efficacy. *Neuropsychopharm* 10:1–7.
- Carlsson M, Carlsson A. 1990. Interactions between glutamatergic and monoaminergic systems within the basal ganglia—implications for schizophrenia and Parkinson's disease. *Trends Neurosci* 13:272–6.
- Carter CJ, LHeureux R, Scatton B. 1988. Differential control by N-methyl-D-aspartate and kainate of striatal dopamine release in vivo: a trans-striatal dialysis study. *J Neurochem* 51:462–8.
- Casey DE. 1993. Serotonergic and dopaminergic aspects of neuroleptic-induced extrapyramidal syndromes in nonhuman primates. *Psychopharm. (Berl)* 112:S55–9.
- Charley PJ, Grenhoff J, Chergui K, De la Chapelle B, Buda M, Svensson TH, Chouvet G. 1991. Burst firing of mesencephalic dopamine neurons is inhibited by somatodendritic application of kynurenic acid. *Acta Physiol Scand* 142:105–12.
- Clow DW, Jhamandas K. 1989. Characterization of L-glutamate action on the release of endogenous dopamine from the rat caudate-putamen. *J Pharmacol Exp Ther* 248:722–8.
- Coughenour LL, Cordon JJ. 1997. Characterization of haloperidol and trifluoperidol as subtype-selective N-methyl-D-aspartate (NMDA) receptor antagonists using [<sup>3</sup>H]TCP and [<sup>3</sup>H]ifenprodil binding in rat brain membranes. *J Pharmacol Exp Ther* 280:584–92.
- Crowder JM, Bradford HF. 1987. Inhibitory effects of noradrenaline and dopamine on calcium influx and neurotransmitter glutamate release in mammalian brain slices. *Eur J Pharmacol* 143:343–52.
- Deakin JF, Slater P, Simpson MD, Gilchrist AC, Skan WJ, Royston MC, Reynolds GP, Cross AJ. 1989. Frontal cortical and left temporal glutamatergic dysfunction in schizophrenia. *J Neurochem* 52:1781–6.
- Deutch AY, Moghaddam B, Innis RB, Krystal JH, Aghajanian GK, Bunney BS, Charney DS. 1991. Mechanisms of action of atypical antipsychotic drugs. *Schizophr Res* 4:121–156.
- Deutch SI, Mastropalo J, Schwartz BL, Rosse RB, Morihisa JM. 1989. A "glutamatergic hypothesis" of schizophrenia. Rationale for pharmacotherapy with glycine. *Clin Neuropharmacol* 12:1–13.
- Elliott PJ, Close SP, Walsh DM, Hayes AG, Marriott AS. 1990. Neuroleptic-induced catalepsy as a model of Parkinson's disease. II. Effect of glutamate antagonists. *J Neural Transm: Park Dis Dement Sect* 2:91–100.
- Farde L, Wiesel FA, Halldin C, Sedvall G. 1988. Central D2-dopamine receptor occupancy in schizophrenic patients treated with antipsychotic drugs. *Arch Gen Psychiatry* 45:71–6.
- Faustman WO, Bardgett M, Faull KF, Pfefferbaum A, Csernansky J. 1999. Cerebrospinal fluid glutamate inversely correlates with positive symptom severity in unmedicated male schizophrenic/schizoaffective patients. *Biol Psychiatry* 45:68–75.
- Fitzgerald LW, Deutch AY, Gasic G, Heinemann SF, Nestler EJ. 1995. Regulation of cortical and subcortical glutamate receptor subunit expression by antipsychotic drugs. *J Neurosci* 15:2453–61.
- Freed WJ. 1988. The therapeutic latency of neuroleptic drugs and nonspecific postjunctional supersensitivity. *Schizophr Bull* 14:269–77.
- Gattaz WF, Gasser T, Beckmann H. 1985. Multidimensional analysis of the concentrations of 17 substances in the CSF of schizophrenics and controls. *Biol Psychiatry* 20:360–6.
- Giardino L, Bortolotti F, Orzazo C, Pozza M, Monteleone P, Calza L, Maj M. 1997. Effect of chronic clozapine administration on [<sup>3</sup>H]MK801-binding sites in the rat brain: a side-preference action in cortical areas. *Brain Res* 762:216–8.
- Goldstein JM, Litwin LC, Sutton EB, Malick JB. 1989. Effects of ICI 169,369, a selective serotonin<sub>2</sub> antagonist, in electrophysiological tests predictive of antipsychotic activity. *J Pharmacol Exp Ther* 249:673–80.

- Hamid EH, Hyde TM, Baca SM, Egan MF. 1998. Failure to down regulate NMDA receptors in the striatum and nucleus accumbens associated with neuroleptic-induced dyskinesia. *Brain Res* 796:291-295.
- Healy DJ, Meador-Woodruff JH. 1997. Clozapine and haloperidol differentially affect AMPA and kainate receptor subunit mRNA levels in rat cortex and striatum. *Brain Res Mol Brain Res* 47:331-8.
- Huttunen M. 1995. The evolution of the serotonin-dopamine antagonist concept. *J Clin Psychopharmacol* 15:4S-10S.
- Imperato A, Honore T, Jensen LH. 1990. Dopamine release in the nucleus caudatus and in the nucleus accumbens is under glutamatergic control through non-NMDA receptors: a study in freely moving rats [published erratum appears in *Brain Res* (1991) 539:179]. *Brain Res* 530:223-8.
- Janowsky A, Berger SP. 1989. Clozapine inhibits [3H] MK-801 binding to the glutamate receptor ion channel complex. *Schizophr Res* 2:189.
- Kakigi T, Gao XM, Shirakawa O, Tamminga CA. 1992. Chronic haloperidol treatment reduces glutamate receptor binding in the limbic structures of rat brain. *Soc Neurosci Abstracts* 18:277.
- Kerwin R. 1994. The new atypical antipsychotics. *Br J Psychiatry* 164:141-148.
- Kerwin R, Patel S, Meldrum B. 1990. Quantitative autoradiographic analysis of glutamate binding sites in the hippocampal formation in normal and schizophrenic brain post mortem. *Neuroscience* 39:25-32.
- Kerwin RW, Patel S, Meldrum BS, Czudek C, Reynolds GP. 1988. Asymmetrical loss of glutamate receptor subtype in left hippocampus in schizophrenia [letter]. *Lancet* 1:583-4.
- Kim JS, Kornhuber HH, Schmid-Burgk W, Holzmüller B. 1980. Low cerebrospinal fluid glutamate in schizophrenic patients and a new hypothesis on schizophrenia. *Neurosci Lett* 20:379-82.
- Kornhuber J, Kornhuber ME. 1986. Presynaptic dopamine modulation of cortical input to the striatum. *Life Sci* 39:669-674.
- Kornhuber J, Mack-Burkhardt F, Riederer P, Hebenstreit GF, Reynolds GP, Andrews HB, Beckmann H. 1989. [3H]MK-801 binding sites in postmortem brain regions of schizophrenic patients. *J Neural Transm* 77:231-6.
- Korpi ER, Kaufmann CA, Marnela KM, Weinberger DR. 1987. Cerebrospinal fluid amino acid concentrations in chronic schizophrenia. *Psychiatry Res* 20:337-45.
- Krebs MO, Desce JM, Kemel ML, Gauchy C, Godeheu G, Chéramy A, Glowinski J. 1991. Glutamatergic control of dopamine release in the rat striatum: evidence for presynaptic N-methyl-D-aspartate receptors on dopaminergic nerve terminals. *J Neurochem* 56:81-5.
- Lidsky TI, Banerjee SP. 1992. Clozapine's mechanisms of action: non-dopaminergic activity rather than anatomical selectivity [published erratum appears in *Neurosci Lett* (1992) 146:119]. *Neurosci Lett* 139:100-3.
- Lidsky TI, Yablonsky-Alter E, Zuck L, Banerjee SP. 1993. Anti-glutamatergic effects of clozapine. *Neurosci Lett* 163:155-8.
- McCoy L, Cox C, Richfield EK. 1998. Antipsychotic drug regulation of AMPA receptor affinity states and GluR1, GluR2 splice variant expression. *Synapse* 28:195-207.
- McMillen BA, Shore PA. 1978. Comparative effects of clozapine and alpha-adrenoceptor blocking drugs on regional noradrenaline metabolism in rat brain. *Eur J Pharmacol* 52:225-30.
- Meltzer HY. 1991. The mechanism of action of novel antipsychotic drugs. *Schizophr Bull* 17:263-87.
- Meltzer HY, Matsubara S, Lee JC. 1989. Classification of typical and atypical antipsychotic drugs on the basis of dopamine D-1, D-2 and serotonin<sub>2</sub> pKi values. *J Pharmacol Exp Ther* 251:238-46.
- Meshul CK, Bunker GL, Mason JN, Allen C, Janowsky A. 1996. Effects of subchronic clozapine and haloperidol on striatal glutamatergic synapses. *J Neurochem* 67:1965-73.
- Mitchell PR, Doggett NS. 1980. Modulation of striatal [3H]-glutamic acid release by dopaminergic drugs. *Life Sci* 26:2073-81.
- Nishikawa T, Takashima M, Toru M. 1983. Increased [3H]kainic acid binding in the prefrontal cortex in schizophrenia. *Neurosci Lett* 40:245-50.
- Noga JT, Hyde TM, Herman MM, Spurney CF, Bigelow LB, Weinberger DR, Kleinman JE. 1997. Glutamate receptors in the postmortem striatum of schizophrenic, suicide, and control brains. *Synapse* 27:168-76.
- Oretti RG, Spurlock G, Buckland PR, McGuffin P. 1994. Lack of effect of antipsychotic and antidepressant drugs on glutamate receptor mRNA levels in rat brains. *Neurosci Lett* 177:39-43.
- Peroutka SJ, Snyder SH. 1980. Relationship of neuroleptic drug effects at brain dopamine, serotonin, alpha-adrenergic, and histamine receptors to clinical potency. *Am J Psychiatry* 137:1518-22.
- Peroutka SJ, U'Prichard DC, Greenberg DA, Snyder SH. 1977. Neuroleptic drug interactions with norepinephrine alpha receptor binding sites in rat brain. *Neuropharm* 16:549-56.
- Perry TL. 1982. Normal cerebrospinal fluid and brain glutamate levels in schizophrenia do not support the hypothesis of glutamatergic neuronal dysfunction. *Neurosci Lett* 28:81-5.
- Riva MA, Tascadda F, Lovati E, Racagni G. 1997. Regulation of NMDA receptor subunit messenger RNA levels in the rat brain following acute and chronic exposure to antipsychotic drugs. *Brain Res Mol Brain Res* 50:136-42.
- Roberts PJ, Sharif NA. 1978. Effects of l-glutamate and related amino acids upon the release of [3H]dopamine from rat striatal slices. *Brain Res* 157:391-5.
- Seeman P. 1980. Brain dopamine receptors. *Pharmacol Rev* 32:229-313.
- Simpson MD, Slater P, Royston MC, Deakin JF. 1992. Regionally selective deficits in uptake sites for glutamate and gamma-aminobutyric acid in the basal ganglia in schizophrenia. *Psychiatry Res* 42:273-82.
- Subramaniam S, McGonigle P. 1991. Quantitative autoradiographic characterization of the binding of (+)-5-methyl-10,11-dihydro-5H-dibenzo[a,d]cyclohepten-5, 10-imine ([3H]MK-801) in rat brain: regional effects of polyamines. *J Pharmacol Exp Ther* 256:811-9.
- Tarazi FI, Florijn WJ, Creese I. 1996. Regulation of ionotropic glutamate receptors following subchronic and chronic treatment with typical and atypical antipsychotics. *Psychopharm (Berl)* 128:371-9.
- Toru M, Watanabe S, Shibuya H, Nishikawa T, Noda K, Mitsushio H, Ichikawa H, Kurumaji A, Takashima M, Mataga N, et al. 1988. Neurotransmitters, receptors and neuropeptides in post-mortem brains of chronic schizophrenic patients. *Acta Psychiatr Scand* 78:121-37.
- Van Tol HH, Bunzow JR, Guan HC, Sunahara RK, Seeman P, Niznik HB, Civelli O. 1991. Cloning of the gene for a human dopamine D4 receptor with high affinity for the antipsychotic clozapine. *Nature* 350:610-4.
- Wachtel H, Turski L. 1990. Glutamate: a new target in schizophrenia? [see comments]. *Trends Pharmacol Sci* 11:219-20.
- Wiesel FA, Nordstrom AL, Farde L, Eriksson B. 1994. An open clinical and biochemical study of ritanserin in acute patients with schizophrenia. *Psychopharm (Berl)* 114:31-8.
- Yamamoto BK, Cooperman MA. 1994. Differential effects of chronic antipsychotic drug treatment on extracellular glutamate and dopamine concentrations. *J Neurosci* 14:4159-66.
- Yamamoto BK, Pehek EA, Meltzer HY. 1994. Brain region effects of clozapine on amino acid and monoamine transmission. *J Clin Psychiatry* 55:Suppl B 8-14.
- Yoshida Y, Ono T, Kizu A, Fukushima R, Miyagishi T. 1991. Striatal N-methyl-D-aspartate receptors in haloperidol-induced catalepsy. *Eur J Pharmacol* 203:173-80.